Name:	**C5 Protein**
Catalog Number:	**A120**
Sizes Available:	250 µg/vial
Concentration:	1.0 mg/mL (see Certificate of Analysis for actual concentration)
Form:	Frozen liquid
Activity:	>80% versus normal human serum standard.
Purity:	>95% by SDS-PAGE
Buffer:	10 mM sodium phosphate, 145 mM NaCl, pH 7.2
Extinction Coeff.	$A_{280\ nm}$ = 1.03 at 1.0 mg/mL
Molecular Weight:	190,000 Da (2 chains)
Preservative:	None, 0.22 µm filtered
Storage:	-70°C or below. Avoid freeze/thaw.
Source:	Normal human serum (shown by certified tests to be negative for HBsAg, HTLV-I/II, STS, and for antibodies to HCV, HIV-1 and HIV-II).
Precautions:	Use normal precautions for handling human blood products.
Origin:	Manufactured in the USA.

General Description

Native human C5 is a naturally glycosylated (1.6%) polypeptide containing two disulfide-linked chains. C5 is essential for formation of the membrane attack complex (MAC) and is activated by all three pathways of complement activation. Each pathway of complement activation generates proteolytic enzyme complexes (C3/C5 convertases) which are bound to the target surface (Ross, G.D. (1986)). These enzymes cleave a peptide bond in the larger alpha chain of C5 releasing the highly potent anaphylatoxin C5a (74 amino acids) and activating C5b. This is the only proteolytic step in the assembly of the C5b-9 complex. C5b is unstable, but it remains bound to the activating complex for a brief time (~2 min) during which time it either binds a single C6 from the surrounding fluid to form C5b,6 or it decays and aggregates and is no longer capable of forming MAC. The C5b,6 complex may also remain bound to the C3/C5 convertase where the binding of a single C7 exposes a membrane-binding region and C5b,6,7 can partially insert into the bilipid layer of the target cell. Up to this point the complex may diffuse away from the target cell and enter the membrane of a nearby cell. This is called bystander lysis or “reactive lysis” and can be a significant source of pathology. Each C5b-7 complex can bind one C8 protein molecule which results in the complex inserting more firmly into the membrane. This complex binds C9 and each bound C9 can bind another C9 initiating formation of a ring structure containing up to 18 C9 molecules (Podack, E.R. (1984)). C5b-9 complexes with one or more C9 are referred to as the Membrane Attack Complex (MAC) of complement. Not all C5b-8 complexes have complete rings of C9 with the average being only three C9 per C5b-8 complex. Completed protein rings of C9 form the pores seen on electron micrographs and they result in leakage of metabolites and small proteins out of the cell as well as movement of water into the cell. If sufficient numbers are inserted into a cell membrane then water flowing into the cell, due to osmotic pressure, will rupture the cell membrane allowing the entire contents of the target cell (or a bystander cell) to be released. Either process may result in cell death. Originally it was thought that this required only one C5b-9 complex per cell (referred to as the “one hit theory” of lysis (Rommel F.A. and Mayer,

M.M. (1973)), but this is probably not correct. For example, an erythrocyte requires approximately 850 C5b-9 complexes, as measured by the number of C7 molecules, for lysis to occur (Bauer, J. et al. (1979)). Host cells protected from MAC by CD59 require sufficient numbers of C5b-9 to tie up all the CD59 and then approximately 850 more C5b-9 in addition. Lysis of nucleated cells requires many more C5b-9 complexes due to their size and due to the presence of multiple defense mechanisms in such cells.

Physical Characteristics & Structure

Molecular weight: 190,000 Da composed of two disulfide linked chains. The alpha chain is 115,000 Da and the beta chain is 75,000 Da. Alpha and beta chains are linked through a disulfide bonds. The pI of C5 is 4.7 to 5.5.

Cleavage of C5 by C3/C5 convertases releases C5a (a 74 amino acid fragment, 8268 Da deglycosylated, 10,400 $\pm$ 1000 glycosylated) from the N-terminal of the alpha chain. The C5b fragment (181,000 Da) binds to C6 to initiate the spontaneous assembly of the membrane attack complex.

CAS Number: 80295-53-0

Function

See General Description above.

Assays

The simplest assay for C5 is to use C5-depleted human serum and measure the lysis of EA (classical pathway) or Er (alternative pathway) as a function of the concentration of added test sample or standard purified C5. Each unique application might require appropriate conditions to be determined. However, a typical assay would involve mixing on wet ice 25 µL C5-Dpl, C5-containing sample diluted with GVB++ to contain from 1 to 10 ng C5, and sufficient GVB++ to bring the volume to 300 µL. EA (3×10^7 cells in 200 µL) diluted in GVB++ should be added last. Purified C5 or normal human serum (NHS) may be used as a source of C5. The reaction mixture is incubated for 30 min at 37ºC and 1 mL of cold GVBE is added. The reaction is then mixed and centrifuged to spin down unlysed cells. The released hemoglobin in the supernatant is then analyzed at 415 nm and compared to blanks without C5 (background lysis control) and cells incubated with 275 µL water in place of GVB++ and 25 µL C5-Dpl (100% lysis control).

Many other assays have been described using EA preloaded with C1 (EAC1 cells) or preloaded with the classical pathway C5 convertase (EAC1423 cells). However, all these assays require the use of multiple purified complement components or more difficult-to-prepare reagents (Dodds, A.W. and Sim, R.B. (1997); Morgan, B.P. (2000); Tack, B.F., et al. (1981)).

Applications

See General Description above.

In vivo

The normal serum concentration of C5 is 75 µg/mL (normal range 55 to 113 µg/mL). The primary site of synthesis is the liver, but C5 is also made in the lung (by type II alveolar cells), spleen, intestine, monocytes, and macrophages. The cytokines that

stimulate increased biosynthesis of many other complement proteins do not affect C5 expression levels. C5 is not an acute-phase protein.

Regulation

Many proteins and other components of plasma have an inhibitory effect on the lytic activity of C5b-9 complexes. Most can interact with the forming complex after the C5b-7 stage If any of the C5b-containing complexes fail to insert into a membrane they may self-aggregate or bind to regulatory proteins the most prevalent of which is S Protein. S Protein (also called vitronectin) is an 80,000 Da plasma protein that binds to C5b-9 complexes that fail to insert in the target cell membrane. This reduces damage to nearby host cells. Many other serum components inhibit or partially inhibit lysis by C5b-9 and these include SP40,40 (also known as clusterin and apolipoprotein J) and many plasma lipoprotein complexes (LDL, HDL, etc.).

There are no C5b inactivators that function like factors H and I for C3b or C4bBP and factor I for C4b. That is, C5b is not converted proteolytically to an iC5b form by any known system.

Host cells protect themselves from C5b-9 by a variety of mechanisms. Membrane proteins DAF, MCP, and CR1 inhibit formation of C3/C5 convertases preventing MAC formation. CD59, also called "homologous restriction factor" and "protectin", is a 18,000 to 20,000 Da ubiquitous component of cell membranes that is very effective at binding to and inhibiting the lytic potential of C5b-8 and C5b-9 complexes. The species-specificity of CD59 is not absolute and many mammalian CD59 does inhibit or partially inhibit MAC from other species. The specificity that is observed appears to be due to incompatibility between the C8 of one animal and the CD59 of another. Like DAF, CD59 contains a GPI anchor (a post-translationally added lipid tail that inserts into the bilipid layer of the cell). The disease PNH is caused by the loss of enzymes that attach the GPI tail, thus depriving cells of the ability to inactivate C3/C5 convertases and the ability to inactivate C5b-9. This results in complement-mediated damage to and eventual lysis of long-lived blood cells such as erythrocytes and platelets.

Biosynthesis of C5 is upregulated in most cells that synthesize C5 by IL-1, IL-6, TNF-alpha, and LPS.

Genetics

Human chromosome location 9q 32-34. Accession number M57729. Mouse chromosome 2. Accession number J05234. Human genomic structure: the gene spans 79 kb with 41 exons.

Deficiencies

A significant number of common mouse strains lack C5 due to loss during an early step in breeding the laboratory inbred mouse. Six C5-deficient strains are common A/HeJ, AKR/J, DBA/2J, NZB/B1NJ, SWR/J, and B10.D2/oSnJ. All lack C5 due to a two base pair deletion near the 5' end of the cDNA. Clearly survival does not depend on C5 or the ability to form MAC, however, these mice have shown many immune system abnormalities and are generally more susceptible to many infections diseases. Similarly, several C5-deficient families have been reported. C5-deficient humans have severely impaired bactericidal and chemotactic activities. Recurrent infections are common and neisserial species are a particularly serious problem.

Diseases

See Deficiencies above.

Precautions/Toxicity/Hazards

This protein is purified from human plasma, therefore precautions appropriate for handling any blood-derived product must be used even though the source was shown by certified tests to be negative for HBsAg, HTLV-I/II, STS, and for antibodies to HCV, HIV-1 and HIV-II.

Hazard Code: B WGK Germany 3

MSDS available upon request.

References

Bauer, J., Podack, E.R. and Valet, G. (1979) Determination of the number of lytic sites in biconcave and spheroid erythrocyte ghosts after complement lysis. J. Immunol. 122:2032-2036.

Dodds, A.W. and Sim, R.B. editors (1997) Complement. A Practical Approach (ISBN 019963539) Oxford University Press, Oxford.

Law, S.K.A. and Reid, K.B.M. (1995) Complement 2nd Edition (ISBN 0199633568) Oxford University Press, Oxford.

Morgan, B.P. ed. (2000) Complement Methods and Protocols. (ISBN 0-89603-654-5) Humana Press, Inc., Totowa, New Jersey.

Morley, B.J. and Walport, M.J. (2000) The Complement Facts Book (ISBN 0127333606) Academic Press, London.

Podack, E.R. (1984) Molecular composition of the tubular structure of the membrane attack complex of complement. J. Biol. Chem. 259: 8641-8647.

Rommel, F.A. and Mayer, M.M. (1973) Studies of guinea pig complement component C9: reaction kinetics and evidence that lysis of EAC1-8 results from a single membrane lesion caused by one molecule of C9. J. Immunol. 110:637-647.

Ross, G.D. (1986) Immunobiology of the Complement System. (ISBN 0-12-5976402) Academic Press, Orlando.

Tack, B.F., Janatova, J., Thomas, M.L., Harrison, R.A. and Hammer, C.H. (1980) The third, fourth and fifth components of human complement: Isolation and biochemical properties. Methods in Enzymology 80, 64-101.

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